



METAGENOMICS AND ITS APPLICATION IN RUMEN ECOSYSTEM: POTENTIAL BIOTECHNOLOGICAL PROSPECTS

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Introduction

- The microbial populations are vital to life on the earth and are of enormous practical significance in medicine; engineering and agriculture (Sloan *et al.*, 2006).

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Microbial ecology seeks to answer 3 questions:

1. Who's there?
2. Who's active?
3. What are they doing?

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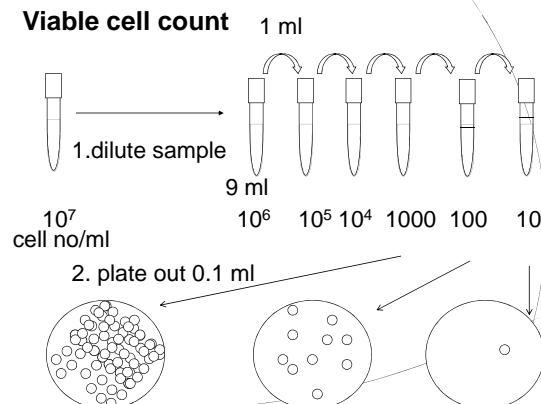


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Viable cell count



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What is metagenomics:

- The term "metagenomics" was first coined by Handelsman *et al.* (1998) to study the genomes from all microbes in a particular environment as opposed to the genome from one organism isolated from the environment and cultured *in vitro*.



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Molecular methods for enumeration

- Genetic-based methods are rapidly replacing conventional detection and enumeration methods in microbiology.
- These methods are mainly based on small subunit ribosomal RNA sequences.

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Why molecular methods?

- The majority of environmental microbes are not easily cultured by usual lab methods.
- Microbes are morphologically indistinct.
- Culture-independent molecular methods can be used to distinguish bacterial populations and to describe microbial communities.
- Most of these methods involve PCR amplification of DNA extracted from natural samples.

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The 16S ribosomal RNA molecule



The 16S rRNA molecule is a major component of the small ribosomal subunit. It has approximately 1500 ribonucleotides. This single-stranded rRNA molecule has an intricate secondary structure with extensive intrachain base pairing. The 16S rRNA forms a part of the ribosomal structure that is the site of protein biosynthesis resulting in the translation of messenger RNA. The 3' end of the bacterial 16S rRNA base-pairs with the Shine-Dalgarno sequence located upstream of the AUG initiation codon in mRNA during the initiation step of the translation process. This allows the mRNA to position itself on the ribosome. There is also evidence that 16S rRNA is directly involved in the interactions between the large and small ribosomal subunits.

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DNA extraction

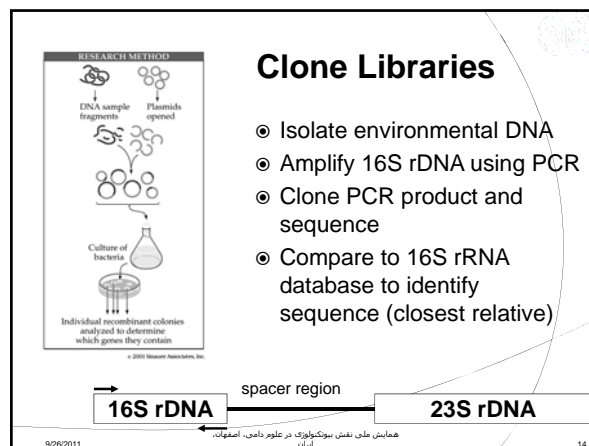
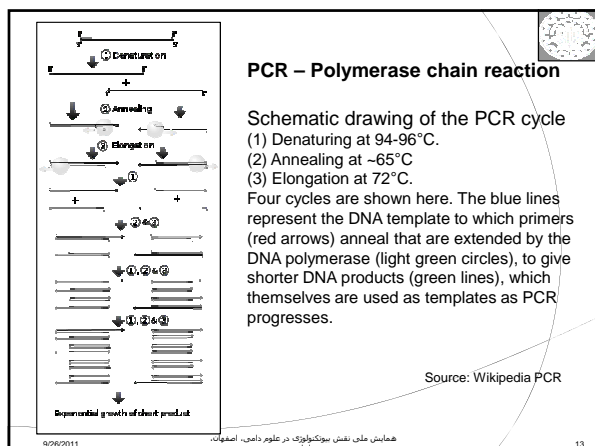
Three steps:

1. **Lysis of the bacterial cells.**
 - mechanical means (physically breaking open the cells)
 - chemical means (using enzymes that break down the bacterial cell walls).
2. **Protecting the DNA from degradation.** Proteins are precipitated, leaving the DNA in solution. This removes enzymes (proteins) that can break down DNA.
3. **Purifying the DNA.** DNA is precipitated with ethanol, and cellular components that do not precipitate with ethanol are washed away. The precipitated DNA is dried and resuspended in water or buffer.

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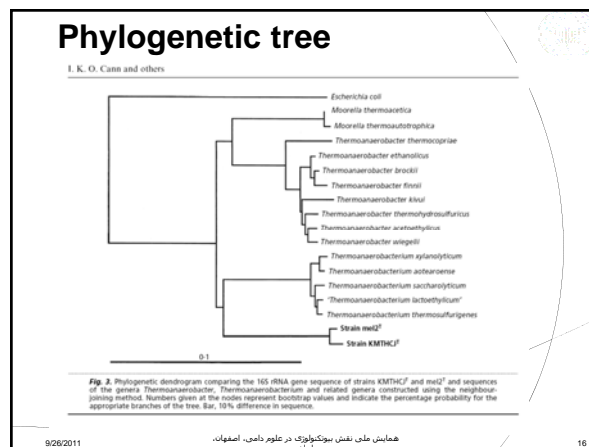


Bacillus sp. ICPS6 16S rDNA

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1 agtttgatcc tggctcagga cgaagctgg cggcgtgct aatacatgca agtcgagcgg
61 acgggggggg agcttcttc cgtctggta cggcgggag ggtgagtaac acgtgggtaa
121 cctgcggata agacgggat aactcggga aacgggggt aatccggat aacacggag
181 aacgcatggt cttcgtgtga aaggcgctt cggctgcaa ttacggaggg gcccgcgcg
241 cattagctag ttgctgggt aacgctcac caaggcgag atgcgtagcc ggccctggag
301 ggtgacggc cacactggga ctgagacac gccagactc ctacgggag cagcagtagg
361 gaattctcgg caatggcga aagcttgac gacggagcc gctgagcga agaagctcct
421 cggatcgtaa agctctgttg ttaggaaag aagagtcgg ttccaaacgg cggcggcgt
481 gacggtaact aacggaagaa ccggcgtaa ctacgtgca gaagccggg taatacgtag
541 gggcgagcg ttgtcggaa ttatggggc taagcgggc gaagcggtc ccttaagtct
601 gatgtgaaag ccacagctt aactgggag ggtcattga aactgggga cttgagtgca
661 gaagcgaga gggagctcc acgtctagc gtaagtggg tagagctgg gaggaaacc
721 agtggcgag cggctctgt ggtctgaac tgacgtgag ggcggaagc gtgggggca
781 aacaggata gatacctgg tagtcacgc cgtaaacgt gactgttag tgttagggg
841 gttaacctt tagtctgta gctacgcgt taagcactc gctggggag tacggcgca
901 aggtgaaac tcaaggaaat tgaaggggg ccgcaaacg ggtgagcat gtggttaat
961 tcgaagcac ggaagaaac ttacaggtc ttgacatcc ctgacaccc tggagacag
1021 ggttctccc cttggggag gacagggtg caggtgtgc atggttgctc tcagctgtg
1081 tctgagagt ttgggttaag tccgcaaac agcgaaccc tggccctag ttgcagcat
1141 tcaagtggg actctaggg gactccggc taagactcg aggaagctg ggtgacgtc
1201 aatcaccat gcccttatg accctggca caacgtgct caatggcg gtacaaagg
1261 ctgcgaacc ggcagggga ggcattcca aaagccgct ctaagtggc attgagct
1321 gcaactagc tgcattgag cggatcgct agtaatcgg gatcagatg ccgcgggaa
1381 taagtcccg ggcctgttac acaagcccg tcaacacag agagcttga acacgggag
1441 tcggtgaggt aacccgaag ggaagcagc ccgcaaggg gggcaagtg ttgggtgaa
1501 tctgtacaaa

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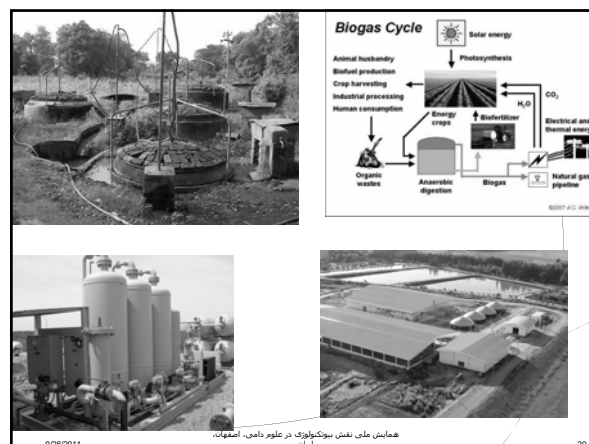
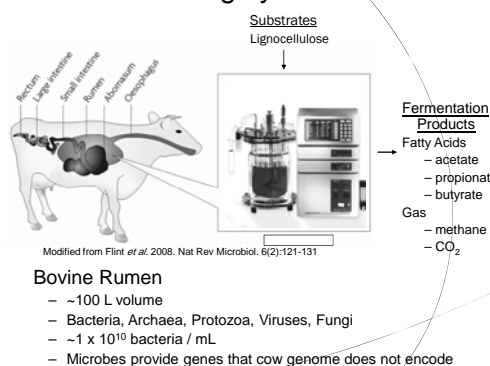
Rumen function and metagenomics:

- Ruminants will probably be important in livestock strategies to assist the poor (Delgado, 2005), therefore their ability to convert locally available feedstuffs to animal products should be improved.
- It's well studied that microbial community inhabiting in the rumen is characterized by its high population density, wide diversity and interactive complexity (Duan *et al.*, 2006).

Rumen function and metagenomics:

- This microbial community is responsible for the bioconversion of lignocellulosic feeds into volatile fatty acids (Kamra, 2005).
- The goal of rumen biotechnologists is to manipulate the ruminal microbial ecosystem to improve the efficiency of feed (Khampa and Wanapat, 2007).

The rumen is a highly efficient bioreactor



Application of rumen metagenomics:

- The development and application of metagenomics has allowed access to the uncultivated ecosystem and insight into metabolic capabilities as yet uncultured microbial communities.
- Some examples of application of metagenomics are as:

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- It is well known that biotechnology has a continuous demand for **novel genes** and enzymes and compounds (Christel *et al.*, of the 2007).
- The rumen microbial diversity represents a vast genetic bounty that may be exploited for the discovery of **novel genes**, **entire metabolic pathways** and potentially **valuable end-products** thereof (Frank and Pace, 2008), but the successful development for the discovery of some novel genes or microbes have not yet been achieved.

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Table 4. The KEGG pathways with significantly different numbers of hits between the two age groups.

KO#	KO pathway (class)	14d ^a	42d ^a	P-value ^b
00230	Purine metabolism (M)	0.0185 ± 1.13E-04	0.0191 ± 3.89E-05	0.0239
00400	Phe, Tyr and Trp biosynthesis (M)	0.0056 ± 1.16E-04	0.0062 ± 7.58E-05	0.0429
03070	Bacterial secretion system (E)	0.0053 ± 3.37E-07	0.0054 ± 1.46E-06	0.0000
00300	Lysine biosynthesis (M)	0.0047 ± 1.85E-05	0.0053 ± 3.59E-05	0.0034
00363	Biosphenol degradation (M)	0.0041 ± 4.59E-06	0.0044 ± 7.24E-05	0.0328
00070	One carbon pool by folate (M)	0.0036 ± 3.06E-05	0.0039 ± 1.13E-05	0.0057
00780	Biotin metabolism (M)	0.0021 ± 6.48E-05	0.0025 ± 5.58E-05	0.0417
00401	Novobiocin biosynthesis (M)	0.0019 ± 4.44E-05	0.0022 ± 2.70E-05	0.0167
03410	Base excision repair (G)	0.0017 ± 1.27E-05	0.0018 ± 2.96E-05	0.0227
00950	Isoquinoline alkaloid biosynthesis (M)	0.0015 ± 1.92E-05	0.0019 ± 5.67E-05	0.0215
00514	O-glycan biosynthesis (M)	0.0001 ± 1.49E-06	0.0002 ± 9.80E-06	0.0192

a. The number of open reading frames (ORF) annotated to a given KO pathway divided by the total number of ORF annotated to all KO pathways (mean ± SEM).

b. P-value was calculated using MetaStats (White *et al.*, 2009).

M, Metabolism; E, Environmental Information Processing; G, Genetic Information Processing.

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Genomic sequences database of the rumen microbes:

- Sequencing the genomes of individual rumen microbes and determining the function of their encoded genes promises to transform our understanding of the microbiology of the rumen (Attwood *et al.*, 2008).

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Table 1: Genome Sequence projects for rumen microorganisms. The institution coordinating the projects and websites for access to data are also shown			
Rumen microbes	Strain and coverage	Institution	websites
Fibrobacter succinogenes	Strain 6 85 closed	TIGR	www.sgr.org/tb/rumenomics
Ruminococcus albus	Strain 8 closed	TIGR	www.sgr.org/tb/rumenomics
Prevotella ruminicola	Strain 23 closed	TIGR	www.sgr.org/tb/rumenomics
Prevotella bryantii	Strain 6, 4, Draft (8x)	TIGR	www.sgr.org/tb/rumenomics
Ruminococcus flavefaciens	Strain FD-1 Draft (2x)	University of Illinois	www.biotech.uiuc.edu

Table 2: Some rumen microbes and enzymes identified using metagenomic tools		
Authors	Enzymes/microbial studies	Source
Ferrer et al., 2005	Novel enzyme RA.04 belonging to the alpha-amylase family	Bovine rumen
Belouqui et al., 2006	Novel gene Ri-5 encoding polyphenol oxidase	Bovine rumen
Mattheu et al., 2007	Novel methanogens	Cattle, Sheep rumen
Lammie et al., 2007	Rumen microbes	Bovine rumen (SSR)
Palackal et al., 2007	multifunctional glycosyl hydrolase	Bovine rumen

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Identification of novel enzymes/microbes from rumen:

- Feral herbivores or the migratory livestock species like goats and sheep could harbor a wealth of valuable GI microorganisms that could be developed and then used as **probiotics or direct fed microbials** (to enhance rumen productivity) and sources of various **hydrolytic enzymes** for promoting livestock nutrition, health and industrial development (Singh *et al.*, 2008) (Table 2).

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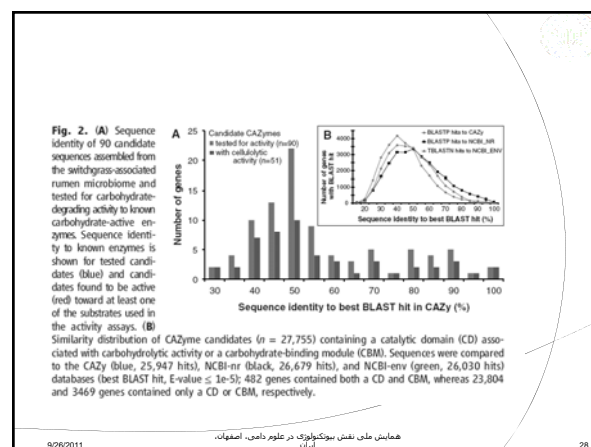


Fig. 1. (A) A surgically created fistula (arrow) sealed with a flexible cannula was used to study the degradation of switchgrass within the rumen. **(B)** Switchgrass before rumen incubation. **(C)** Nylon bags filled with switchgrass before insertion into the rumen. **(D)** Switchgrass after 72 hours of rumen incubation.

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Identification of uncultured methanogens:

- Interest in methanogens from ruminants has resulted from the role of methane from the fact that cattle lose 6% of ingestion energy as methane (Johnson and Johnson, 1995).

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Differentiating and quantitative determination rumen biomass:

- Another important application of microbial metagenomics in animal nutrition is the quantitative determination of total rumen microbial biomass and differentiating the bacterial and protozoal biomass.

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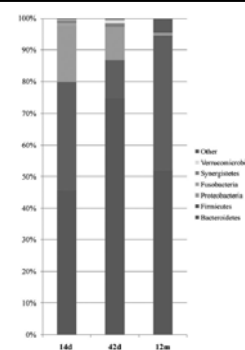


Fig. 1. The relative distribution of the six most abundant phyla in the rumen microbiota of pre-weaned calves fed milk replacer (n=3). Percentages denote numbers of 16S sequences assigned to a given phylum divided by the total number of 16S sequences that were positively assigned to any phylum. The microbial composition of the rumen microbiota of 12-month-old (12m) bull calves (n=4) fed a hay diet was used as a reference.

Environmental Microbiology (2011)
Metagenomics and the rumen microbiota

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Ruminal nitrogen metabolism:

- A better understanding of mechanistic process altering the production and uptake of amino nitrogen will help the livestock nutritionists to improve the overall conversion of dietary nitrogen into microbial protein.

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Conclusion:

- These technologies have the potential to revolutionize the understanding of rumen function and will overcome the limitations of classical based techniques, including isolation and taxonomic identification of strains important to efficient rumen function and better understanding of the roles of microorganisms in relation to achieving high productivity and decreasing environmental pollutants.

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What is metagenomics:

- In principle, any study that addresses all the individuals in a microbial community as a single genomic pool can be seen as an exercise in metagenomics (Kowalchuk *et al.*, 2007).

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- Initially, noncultured microflora and ancient DNA investigations had been the prime targets of metagenomic studies, but presently the technology has been applied to study an array of microbial diversities

like:

deep-sea aquatic microflora,
soil microbes and

GI ecosystem of human and animals (Lu *et al.*, 2007; Shanks *et al.*, 2006).

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Metagenome technologies, DNA extraction, library construction, screening:

- Metagenome analyses are usually initiated by the isolation of environmental DNAs.
- A major difficulty associated with the metagenome approach is related to the contamination of purified DNA with polyphenolic compounds, which are copurified with the DNA.

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Identification of novel enzymes/microbes from rumen:

- The metagenomics need to be exploited to screen and identify novel microbes and biomolecules from the GI tract of the livestock ruminants adapted to the forages or diets enriched with high fiber and an array of anti-nutritional Plant Secondary Metabolites (PSMs) such as tannin-polyphenols.

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